

ISOLATION OF FLAVASPIDIC ACID-PB FROM *DRYOPTERIS SIEBOLDII*

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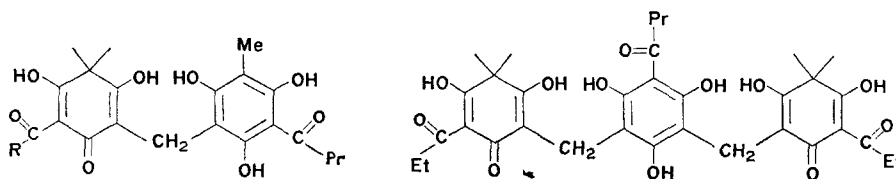
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**Key Word Index**—*Dryopteris sieboldii*; Aspidiaceae; ferns; acylphloroglucinols; flavaspidic acid-AB and -PB; filixic acid -PBP.

**Plant.** *Dryopteris sieboldii* (van Houtte) O. Ktze. **Source.** Kagoshima Prefecture, Japan. **Previous work.** The presence of flavaspidic acid and filixic acid was detected by paper electrophoresis,<sup>1</sup> and the existence of flavaspidic acid-PB was only reported in *D. filix-mas* by PPC.<sup>2</sup>

**Present work.** Dried rhizomes of *D. sieboldii* were percolated with Et<sub>2</sub>O and crude filicin was obtained by MgO method.<sup>3</sup> The Et<sub>2</sub>O solution of crude filicin gave flavaspidic acid-AB (I). Mother liquor after removal of I was chromatographed on silica and eluted with cyclohexane-CHCl<sub>3</sub> (1:1). The elution afforded filixic acid-PBP (II) and then flavaspidic acid-PB (III).



Flavaspidic acid-AB (I) R = Me  
Flavaspidic acid-PB (III) R = Et

Filixic acid-PBP (II)

Flavaspidic acid-AB (I). C<sub>22</sub>H<sub>26</sub>O<sub>8</sub>, m.p. 205–7° (from C<sub>6</sub>H<sub>6</sub>) IR, UV, NMR and m.m.p. with authentic sample. Filixic acid-PBP (II). C<sub>30</sub>H<sub>40</sub>O<sub>12</sub>, m.p. 192–194° (light yellow needles from acetone). IR  $\nu_{\max}^{\text{KBr}}$  cm<sup>-1</sup> 3140 (OH), 2940 (methylene), 1640–1610 (enolic 1,3-diketo system or 2-hydroxyarylyketone), 1192. UV  $\lambda_{\max}^{\text{EtOH}}$  nm (log  $\epsilon$ ) 225 (4.42), 297 (4.24), 345 (4.12). UV  $\lambda_{\max}^{\text{EtOH}+\text{NaOH}}$  nm (log  $\epsilon$ ) 242 (4.37), 315 (4.26). *R<sub>f</sub>* 0.75 on TLC in CHCl<sub>3</sub>-MeOH-H<sub>2</sub>O (7:3:1, lower) spot color gave orange yellow with diazotized benzidine and dark brown with FeCl<sub>3</sub>. The NMR spectrum (NMR analysis in CDCl<sub>3</sub> using TMS as internal reference showed ppm) shows signal attributable to: 0.96 (3H, *t*-COCH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 1.12 (6H, *t*-COCH<sub>2</sub>CH<sub>3</sub>), 1.40, 1.46 (12H, each *s* gem-dimethyl), about 1.76 (2H, *m*-COCH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 3.15 (6H, *t*-COCH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>-COCH<sub>2</sub>CH<sub>3</sub>), 3.52 (4H, *s* two methylene bridges), 9.97 (2H, *s*), 11.39 (1H, *s*), 12.62 (1H, *s*), 15.57 (1H, *s*), 17.78 (2H, *s*), all due to hydrogen bonded hydroxy groups and quenched by addition of D<sub>2</sub>O. The MS, 640 (M<sup>+</sup>), significant peaks at *m/e* 418, 222, 210, 193, 181, 165, 153.

<sup>1</sup> HISADA, S. and NORO, Y. (1961) *Yakugaku Zasshi* **81**, 1270.

<sup>2</sup> PENTILLÄ, A. and SUNDMAN, J. (1964) *Acta Chem. Scand.* **18**, 344.

<sup>3</sup> AEBI, A., BÜCHI, J. and KAPOOR, A. (1957) *Helv. Chim. Acta* **40**, 266.

Flavaspidic acid-*PB* (III).  $C_{23}H_{28}O_8$ , m.p. 154–6° (yellow powder from *n*-hexane). IR  $\nu_{\max}^{KBr}$   $cm^{-1}$  3520, 3230 (OH), 2960 (methylene), 1640–1610 (enolic 1,3-diketo system or 2-hydroxyarylyketone), 1195, 1153. UV  $\lambda_{\max}^{EtOH}$  nm (log  $\epsilon$ ) 223 (4.94), 299 (4.79), 349 (4.93). UV  $\lambda_{\max}^{EtOH+NaOH}$  nm (log  $\epsilon$ ) 239 (4.79), 317 (5.00).  $R_f$  0.14 on TLC in  $CHCl_3$ -MeOH- $H_2O$  (7:3:1, lower) spot color gave orange-red with diazotized benzidine and dark brown with  $FeCl_3$ . The NMR spectrum (NMR analysis in pyridine- $d_5$  using TMS as internal reference showed ppm) shows signal attributable to: 0.90 (3H, *t* -COCH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 1.25 (3H, *t* -COCH<sub>2</sub>CH<sub>3</sub>), 1.56 (6H, *s* *gem*-dimethyl), about 1.85 (2H, *m* -COCH<sub>2</sub>CH<sub>2</sub>Me), 2.38 (3H, *s* aromatic CH<sub>3</sub>), 3.26 (4H, *m* -COCH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>, -COCH<sub>2</sub>CH<sub>3</sub>), 3.99 (2H, *s* methylene bridge between propionylflicinic acid and methylphlorobutyrophenone), 9.33 (5H, *br* OH) quenched by addition of D<sub>2</sub>O. The MS, *m/e* 432 ( $M^+$ ), significant peaks at *m/e* 222, 210, 193, 167.

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## BIFLAVONES IN THE LEAVES OF TWO *JUNIPERUS* PLANTS

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**Key Word Index**—*Juniperus horizontalis*; *Juniperus recurva*; Cupressaceae: biflavones; sciadopitysin; 7,7''-di-*O*-methylcupressuflavone

A number of *Juniperus* plants has already been examined<sup>1–3</sup> for biflavones. We now report the isolation of sciadopitysin<sup>4</sup> (I, 7,4',4'''-tri-*O*-methylamentoflavone) from *Juniperus horizontalis* Moench<sup>5</sup> and 7,7''-di-*O*-methylcupressuflavone<sup>6,7</sup> (II) from *J. recurva* Buch.-Ham.<sup>5</sup> Some other biflavones including cupressuflavone and its monomethyl ether are detected in the leaf extracts of the two plants by TLC examinations. This constitutes the

<sup>1</sup> SAWADA, T., (1958) *J. Pharm. Soc. (Japan)* **78**, 1023; KARIYONE, T. (1962) *J. Pharmacog. Soc. (Japan)* **16**, 1.

<sup>2</sup> MASHIMA, T., OKIGAWA, M. and KAWANO, N. (1970) *J. Pharm. Soc. (Japan)* **90**, 512.

<sup>3</sup> PELTER, A., WARREN, R., HAMEED, N., ILYAS, M. and RAHMAN, W. (1971) *J. Indian Chem Soc* **48**, 204.

<sup>4</sup> KAWANO, N. (1959) *Chem. Pharm. Bull. (Tokyo)* **7**, 698, 821.

<sup>5</sup> Place of collection and identification: Lyod Botanical Garden, Darjeeling, India.

<sup>6</sup> MASHIMA, T., OKIGAWA, M., KAWANO, N., KHAN, N. U., ILYAS, M. and RAHMAN, W. (1970) *Tetrahedron Letters* 2937.

<sup>7</sup> KHAN, N. U., ILYAS, M., RAHMAN, W., MASHIMA, T., OKIGAWA, M. and KAWANO, N. (1973) *Tetrahedron* **29**, in press.